

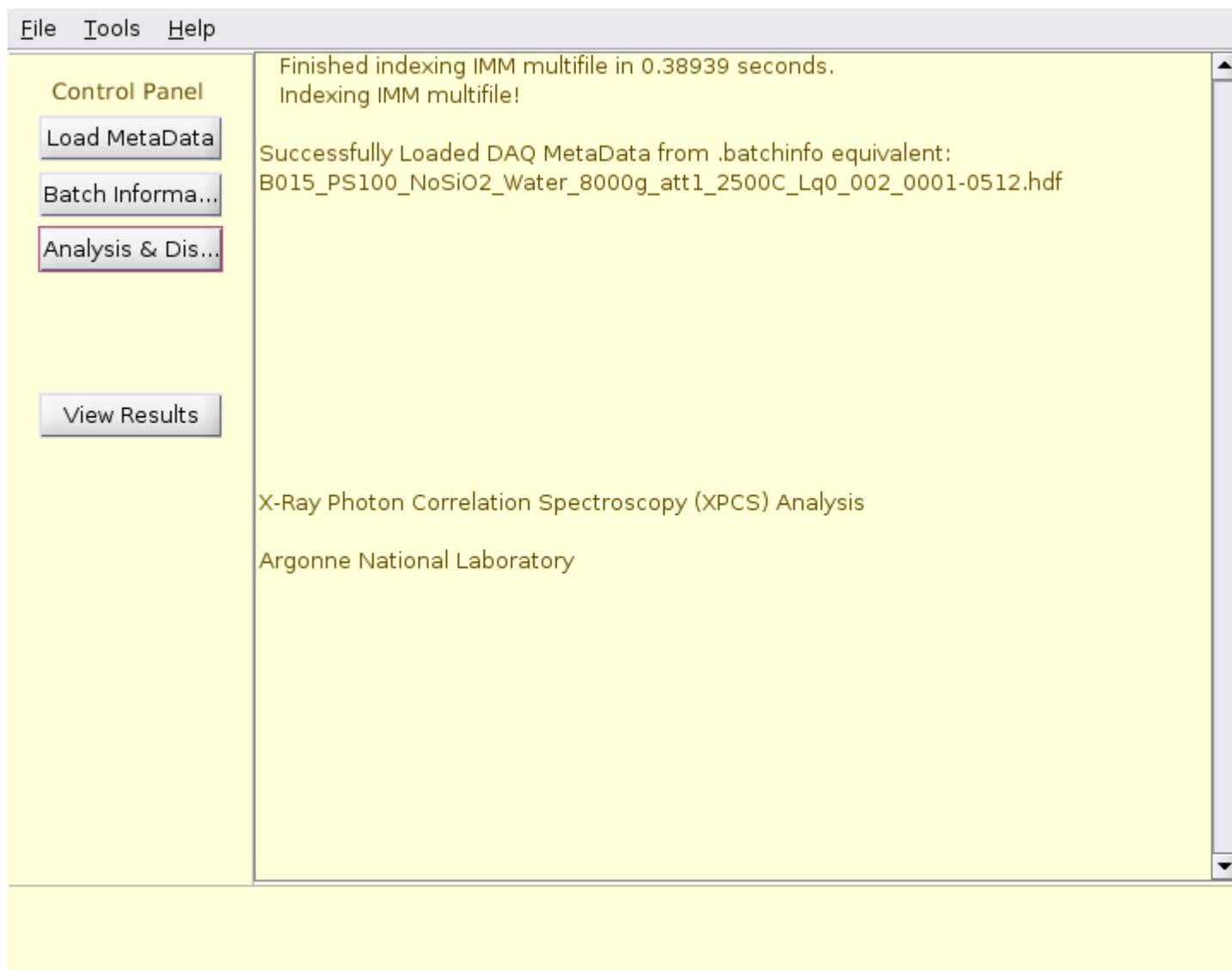
XPCSGUI2 Analysis Procedure Documentation

%%

Modification history:

V1: January 04, 2018

%%



Directory structure of Software:

```
snow% cd /local/XPCSGUI2/
/local/XPCSGUI2
snow% ls
```

```
how_to_set_path.txt      XPCS_DEVELOPMENT      XPCS_LocalProcessing
keep.m                   XPCS_DisplayFiles      XPCS_MetaDataFiles
readme.txt               XPCS_FileReader        XPCS_ProcessingTemplates
set_xpcsgui2_path.m     XPCS_Geometry          XPCS_ToBeDeleted
XPCS_ClusterProcessing  XPCS_GUIFiles          XPCS_Tools
XPCS_DetectorFiles     XPCS_HDF5Files        XPCS_TwoTimeG2
```

The file `how_to_set_path.txt` provides documentation on how to set the path for this software in matlab for your own computer.

Basically, it describes how to edit the file `set_xpcsgui2_path.m` and execute to add the above folders to the path.

The path has to be saved in matlab after this step so that it will remember after the next restart of matlab.

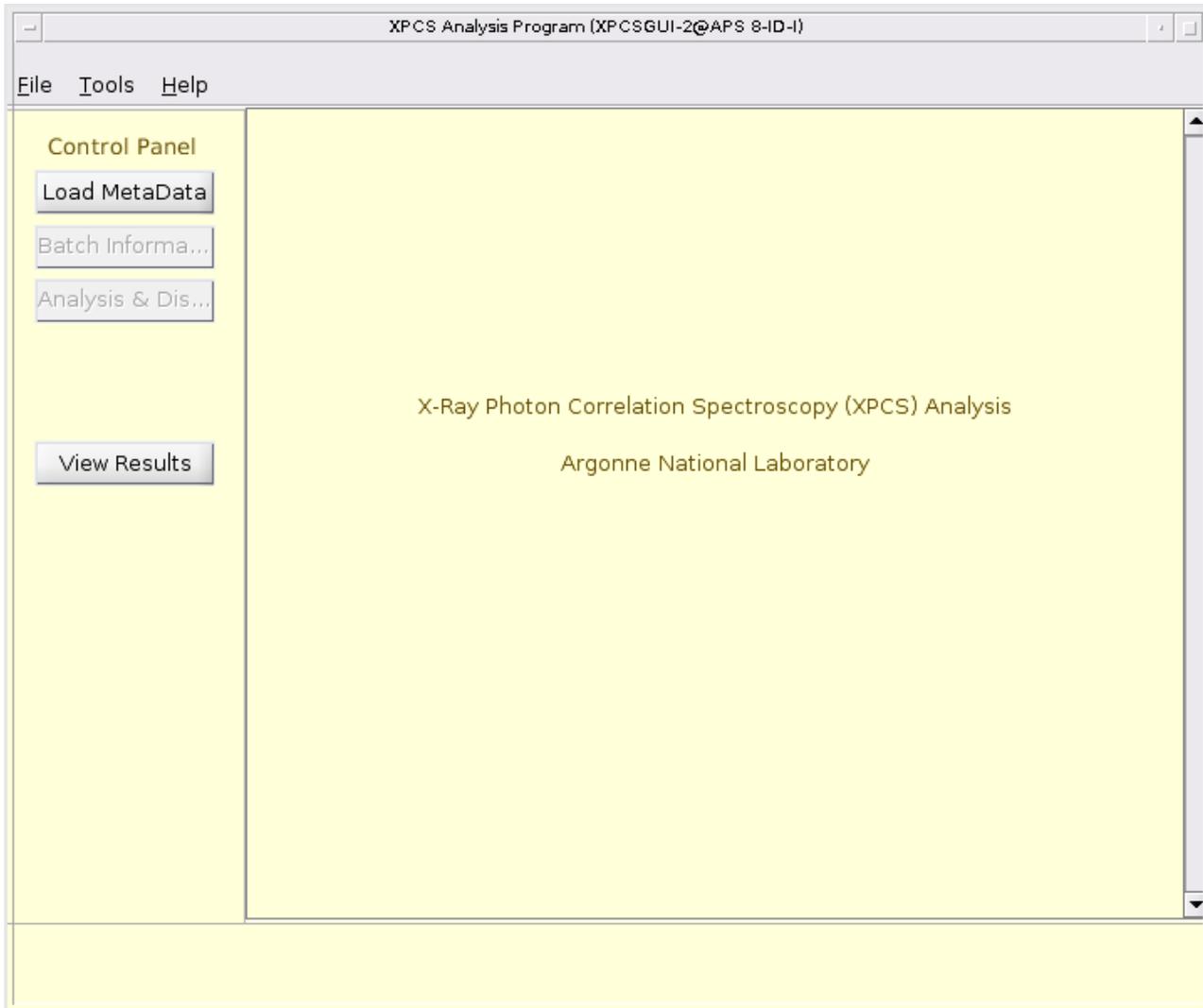
The description also involves editing the file `loadMetaData.m` in the sub-folder `XPCS_MetaDataFiles` to define some defaults for the user computer.

This is shown below for clarity.

```
%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%
%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%
```

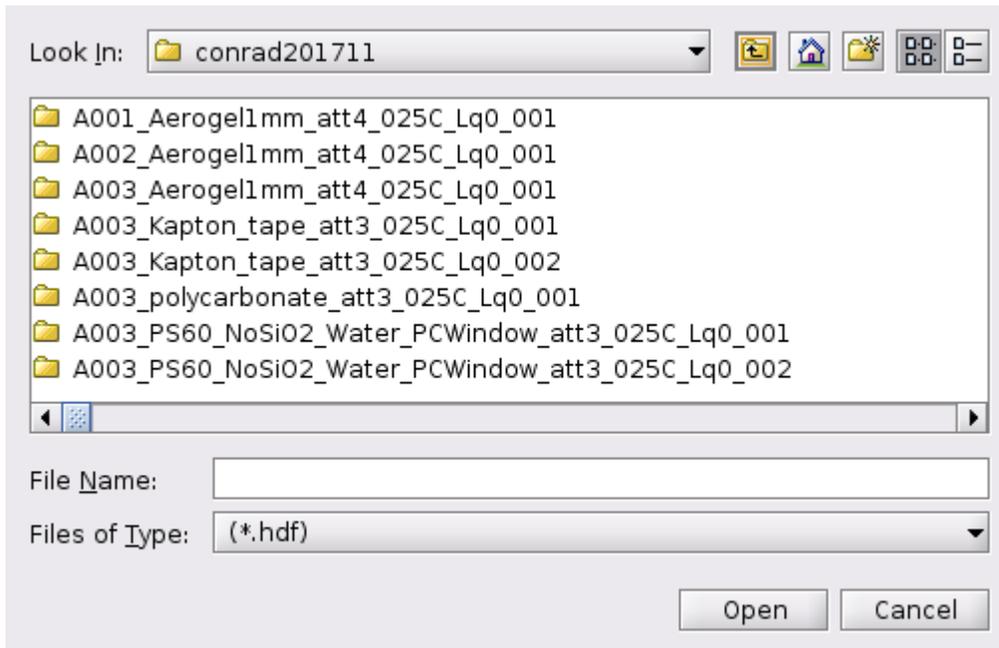
```
ccdimginfo.map_local_location = '/home/8-id-i/partitionMapLibrary/2017-2/';
ccdimginfo.combined_result_folder = 'cluster_results!';
ccdimginfo.force_save_to_matfile=0; %1 will save result as .mat, 0 as .hdf
ccdimginfo.static_mean_window_size = 10; %makes this many parts of the time series for SAXS
(stability
plot)
ccdimginfo.G2_Tensor_Save = 0; % setting to 1 will save the tensors, takes a lot of space
```

```
%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%
%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%%
```



Click on **Load MetaData**

Traverse to the main user data folder (typically PIIastnameYYYYMM) - not the cluster_results folder
pick the desired folder and select the .hdf file (metadata in it, no raw data)



Click on **Batch Information** (usually not much to do in this GUI except for thin films reflection geometry)

In the reflection geometry, the specular direction can be corrected for small errors.

Click on **Show Image** to see a preview of the scattering (usually 100 or so images are averaged)

If any changes are made, click **Apply** and **Close**, else just simply **Close**

Batch Information

Sample-Detector Distance (mm):

Beam Zero Parameters:

x-coordinate in full frame mode

y-coordinate in full frame mode

ccd_{x0} position in beam0 measurer

ccd_{z0} position in beam0 measurer

Specularly Reflected Beam Parameters:

x-coordinate in full frame mode

y-coordinate in full frame mode

ccd_{xspec} in specular measurement

ccd_{zspec} in specular measurement

Saved CCD Image Size:

	column (x)	row (y)
begin	<input type="text" value="0"/>	<input type="text" value="0"/>
end	<input type="text" value="1555"/>	<input type="text" value="515"/>

CCD Position During Data Collection:

ccd_x ccd_z

Experiment Setup Geometry:

X-Ray Photon Energy (KeV):

Nominal Angle:

CCD Working Mode:

kinetics window size

top row number of visible slice

first usable kinetics slice

last usable kinetics slice

View and Edit Batch: of 1 Batches:

	start	end	time (sec)
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data	<input type="text" value="1"/>	<input type="text" value="512"/>	<input type="text" value="0.1"/>
------	--------------------------------	----------------------------------	----------------------------------

dark	<input type="text" value="-1"/>	<input type="text" value="-1"/>	<input type="text" value="0.1"/>
------	---------------------------------	---------------------------------	----------------------------------

Trans. flux (photons/sec)

current (mA)

Batches To Be Analyzed:

Software Pixel Binning

of x pixels to bin (column)

of y pixels to bin (row)

Stride/Skip Frames

of Frames to Stride/Skip

Data Compression:

Status: Ready!

Close

Apply

Show Image

Click on **Analysis & Dis..** (Analysis & Display Settings)

This is the main part.

Analysis Settings

Select Analysis Type:

- static analysis
 dynamic analysis

Partition Map Variable:

Map Map

Static Partition Map 1:

method
Number

Static Partition Map 2:

method
Number

Dynamic Partition Map 1:

method
Number

Dynamic Partition Map 2:

method
Number

Mask Settings:

Default: use the full image

Dynamic Analysis Options:

of delays per multiple tau level

Lower Level Discrimination (LLD):

no LLD X ADU X dark RMS

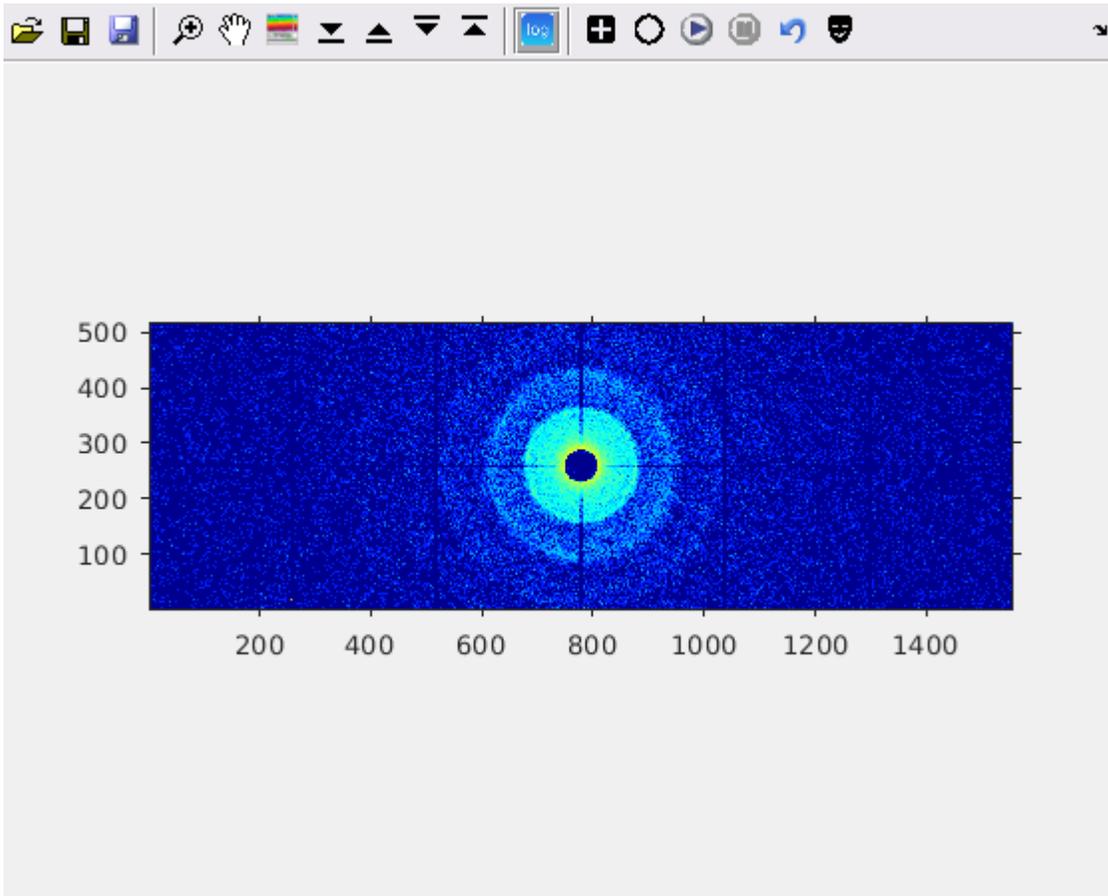
SAVE q/phi digitized pixel map to a .h5 FILE

Enter q/phi map name:

q/phi Map Location:

Status: Ready!

Click on **Mask Polygon** to select a mask (region) to analyze



There are a lot of features in the toolbar icons:

1. "log" will toggle the scale of the image between linear and log.
2. The "+" sign will toggle between + and - which means to include/exclude the region within respectively.
3. The 4 arrow marks are as follows: 2 for raising/lowering the max scale, 2 for raising/lowering the min scale.
4. Or one could click on the colormap next to "Pan" to manually define the color scale for the image.
5. **Open** is to load an existing mask file (.mat), one could load an existing mask from a file and add or remove regions. This is convenient to remove streaks from the scattering that are more random on some sample spots.
6. **Circle** icon will let you draw a circle by clicking on 3 (min) or more points along a best guess circumference. This can be combined with the + and -.
7. Start masking by clicking on the **Play** button and end by clicking on **Pause**. You can continue to make contiguous or disjointed regions by combining **Play** and **Pause** after each block of clicks. Zoom can be combined with drawing mask to seek more clarity.
8. At any point, the **Clear** icon (2nd right) will erase and you can start over with no issue.
9. When finished, it is time to save the mask. This has 2 steps.
10. Left **Save** button saves the mask as a logical matrix to a .mat file with 1 for included pixels and 0 for excluded pixels. This step is optional but highly recommended.
11. Right **Save** button saves the mask matrix to the workspace and **this is a must else the mask will not be included in the qmap and the entire procedure is a waste.**
12. The **mask icon** at the right end shows the mask you have drawn and is good to check.

Show Blemish will display the known bad regions which come from the software database. This gets automatically included on top of the drawn mask.

Selecting Q/Phi Partitions for the Analysis:

This is not described exhaustively below. I would rely on the staff describing the concepts rather than a bunch of lines describing it.

But, basically, there are 2 Maps, naively one is for Static and the other is for Dynamics.

For transmission, (q and phi) are the maps used.

For reflection, (qr and qz) or (qy and qz) are typically used.

It is a MUST to select the ratio of (# in Map 1 / # in Map 2) to be an integer for both Static and Dynamics.

Typical thumb rule is to pick this integer to be some where between 5 and 10.

Note: The spacing in q/phi can be selected to be linear/log as well, will help with coarser binning at higher q where signal is weaker.

Concept:

During correlation analysis, individual pixels are correlated with itself as a function of time. Let us call this as G2 and is of the size of the image array.

Then during the **first normalization step**, G2 from the pixels that belong to the Static partition (Map 1) are averaged together and normalized by the time averaged intensities

over the group of pixels in Map 1. Please note that each bin in Map 2 comprises of integral number of bins in Map 1. In the **second normalization step**, the sub-averages from the first step are further averaged to give you the final g2 as specified in Map 2.

Note: It is also possible and useful in some cases to pick q/phi anisotropic bins for Map 1 and isotropic q/phi=1 bins for Map 2. One example is a case where the structure might be anisotropic and the dynamics could be isotropic.

Saving Qmaps:

On the right side of the Analysis & Display Settings screen, a place to enter the qmap name is provided.

Users are encouraged to follow a standard as the qmaps are saved in a common location for all users.

Suggested template: Pllastname201801_qmap_*****_Lq0_****

PI last name and year and cycle (which should be the main user data folder) and qmap is highly recommended.

The q location is another useful part to have in the name to quickly identify.

Users are welcome to add their sample details and other things as they find it to be helpful to them.

The location where it gets saved is shown below (highlighted in yellow) and this comes from software initialization (loadMetaData.m)

Click on Apply and then Click on "Save map to .h5 file" (which is a .hdf file)

Once it is done, you can click on "View map from .h5 file" to see the qmap.

Note: It is a good idea to view the map to make sure the mask was taken into consideration for example and to check for other aspects.

Running the job:

The GUI functionality stops here as the job running is handled by the scripts.

Typically, there are 3 files that handle the analysis of multitau and two time:

conrad201711_batchmode_XPCS_Job_Creation.m (multitau on the cluster)

Multitau_LocalProcessing_hdf_Template.m (multitau on the local machine)

Twotime_hdf_Template.m (two time on the local machine)